WILEY-VCH

Edited by Ganapathy Subramanian

Continuous Biomanufacturing

Innovative Technologies and Methods

Edited by Ganapathy Subramanian

Continuous Biomanufacturing

Continuous Biomanufacturing

Innovative Technologies and Methods

Edited by Ganapathy Subramanian



Editor

Prof. Dr. Ganapathy Subramanian 44 Oaken Grove SL6 6HH Maidenhead, Berkshire United Kingdom

Cover

Pictures being used: Coagulation factor VIII protein rendering © Molekuul.be; Vitaminpills © cst21; Measurement device containing closed vials © eGraphia. All books published by **Wiley-VCH** are carefully produced. Nevertheless, authors, editors, and publisher do not warrant the information contained in these books, including this book, to be free of errors. Readers are advised to keep in mind that statements, data, illustrations, procedural details or other items may inadvertently be inaccurate.

Library of Congress Card No.: applied for

British Library Cataloguing-in-Publication Data A catalogue record for this book is available from the British Library.

Bibliographic information published by the Deutsche Nationalbibliothek

The Deutsche Nationalbibliothek lists this publication in the Deutsche Nationalbibliografie; detailed bibliographic data are available on the Internet at <<u>http://dnb.d-nb.de</u>/>.

© 2018 Wiley-VCH Verlag GmbH & Co. KGaA, Boschstr. 12, 69469 Weinheim, Germany

All rights reserved (including those of translation into other languages). No part of this book may be reproduced in any form – by photoprinting, microfilm, or any other means – nor transmitted or translated into a machine language without written permission from the publishers. Registered names, trademarks, etc. used in this book, even when not specifically marked as such, are not to be considered unprotected by law.

Print ISBN: 978-3-527-34063-7 ePDF ISBN: 978-3-527-69989-6 ePub ISBN: 978-3-527-69991-9 Mobi ISBN: 978-3-527-69992-6 oBook ISBN: 978-3-527-69990-2

Cover Design Bluesea Design, MacLeese Lake, Canada Typesetting Thomson Digital, Noida, India Printing and Binding

Printed on acid-free paper

 $10 \quad 9 \quad 8 \quad 7 \quad 6 \quad 5 \quad 4 \quad 3 \quad 2 \quad 1$

Contents

List of Contributors *xix*

Part One: Overview of State-of-the-Art Technologies and Challenges 1

v

1	Continuous Bioprocess Development: Methods for Control and		
•	Characterization of the Biological System 3		
	Peter Neubauer and M. Nicolas Cruz-Bournazou		
1.1	Proposed Advantages of Continuous Bioprocessing 3		
1.1.1	Introduction 3		
1.1.1	Special Challenges for Continuous Bioprocesses 5		
1.2	The Biological System in Continuous Biomanufacturing 5		
1.2.1	Inherent Changes in the Microbial System — Droblem of Evolution — 6		
1.2.2	Lack of Process Information 7		
1.2.3.1	Models-Based Process Development and Control for Continuous		
	Processes 8		
1.2.3.2	Engineering Approach to Complex Systems 8		
1.2.4	Limited Control Strategies 9		
1.2.4.1	Traditional Control Strategies for Continuous Cultures 9		
1.3	Changes Required to Integrate Continuous Processes in		
	Biotech 11		
1.3.1	A Better Physiological Understanding of the Organisms and Their		
	Responses on the Reactor Environment 11		
1.3.1.1	Model Complexity 11		
1.3.1.2	Models 12		
1.3.2	Model-Based Process Monitoring 13		
1.3.3	Implementation of Model Predictive Control 14		
1.3.3.1	Model-Based Control 14		
1.4	Role of Iterative Process Development to Push Continuous Processes		
	in Biotech 14		
1.4.1	Methods for Development of Continuous Processes 14		
1.4.1.1	Alternative: Fed-Batch as a System to Simulate Quasi Steady-State		
	Conditions 16		

vi Contents

- 1.4.2 Mimicking Industrial Scale Conditions in the Lab: Continuous-Like Experiments *17*
- 1.4.2.1 A Simple Model for Continuous Processes 17
- 1.4.2.2 Continuous-Like Fed-Batch Cultivations 18
- 1.4.3 Fast and Parallel Experimental Approaches with High Information Content 20
- 1.4.3.1 Computer-Aided Operation of Robotic Facilities 20
- 1.4.3.2 Model Building and Experimental Validation 21
- 1.5 Conclusions 22 References 22
- 2 Tools Enabling Continuous and Integrated Upstream and Downstream Processes in the Manufacturing of Biologicals 31 Rimenvs J. Carvalho and Leda R. Castilho
- 2.1 Introduction *31*
- 2.2 Continuous Upstream Processes 32
- 2.2.1 Continuous Bioprocesses: With or Without Cell Recycle? 33
- 2.2.2 Early/Scale-Down Perfusion Development 34
- 2.2.3 Feeding and Operational Strategies in Perfusion Processes 35
- 2.2.4 Cell Retention Devices 36
- 2.3 Continuous Downstream Processes 41
- 2.3.1 Continuous Liquid Chromatography (CLC) 42
- 2.3.1.1 Continuous Annular Chromatography (CAC) 42
- 2.3.1.2 True and Simulated Moving Bed Chromatography (TMB/SMB) 43
- 2.3.1.3 Multicolumn Countercurrent Solvent Gradient Purification (MCSGP) 45
- 2.3.1.4 Periodic Countercurrent Chromatography (PCC) 47
- 2.3.1.5 Continuous Countercurrent Tangential Chromatography (CCTC) 50
- 2.3.2 Nonchromatographic Continuous Processes 51
- 2.3.2.1 Continuous Aqueous Two-Phase Systems 51
- 2.3.2.2 Continuous Protein Precipitation 52
- 2.3.3 Straight-Through Processes 53
- 2.3.4 Continuous Virus Clearance Processes 54
- 2.4 Integrated Continuous Processes 55
- 2.5 Concluding Remarks 59 References 60

3 Engineering Challenges of Continuous Biomanufacturing Processes (CBP) 69 Holger Thiess, Steffen Zobel-Roos, Petra Gronemeyer, Reinhard Ditz, and Jochen Strube

- 3.1 Introduction 69
- 3.1.1 Continuous Manufacturing 69
- 3.1.2 Continuous Manufacturing of Synthetic Molecules 69
- 3.1.3 Continuous Manufacturing of Biologics 69
- 3.2 Analysis of CBP Status 71

Contents vii

- 3.3 Case Studies 74
- 3.4 Status and Needs for Research and Development 77
- 3.5 Engineering Challenges 79
- 3.5.1 Platform Method of QbD-Driven Process Modeling Instead of Unit Operation Oriented Platform Approaches 80
- 3.5.2 Data Driven Decisions 81
- 3.5.3 Analytics 82
- 3.5.4 QbD Methods 82
- 3.5.5 Upstream and Downstream Integration 82
- 3.5.6 Buffer Handling/Recycling 83
- 3.5.7 Process Integration of Innovative Unit Operations 84
- 3.5.8 ABC (Anything But or Beyond Chromatography) and AAC (Anything and Chromatography) 84
- 3.5.8.1 Liquid–Liquid Extraction Based on ATPE 84
- 3.5.8.2 Precipitation 86
- 3.5.8.3 Membrane Adsorbers 87
- 3.5.8.4 Innovative Materials Like Fibers or Matrices 88
- 3.5.9 Process Concepts for mAbs and Fragments 88
- 3.5.10 Single-Use Technology 91
- 3.5.11 Guided Decision for CBP 91
- 3.6 Conclusion and Outlook 96 Acknowledgments 97 References 97

Part Two: Automation and Monitoring (PAT) 107

4 Progress Toward Automated Single-Use Continuous Monoclonal Antibody Manufacturing via the Protein Refinery Operations Lab 109

David Pollard, Mark Brower, and Douglas Richardson

- 4.1 Introduction 109
- 4.2 Protein Refinery Operations Lab 111
- 4.2.1 Introduction 111
- 4.2.2 Protein Refinery Operations Lab: Design and Implementation 112
- 4.2.3 Protein Refinery Operations Lab: Process Analytical Technology (PAT) and Product Attribute Control (PAC) for the Transition to Real-Time Release (RTR) *117*
- 4.2.3.1 Protein Refinery Operations Lab: Current State of PAT Technologies *118*
- 4.3 Protein Refinery Operations Lab: Case Studies 122
- 4.3.1 Case Study: Perfusion 122
- 4.3.2 Case Study: Continuous Purification 124
- 4.3.3 Case Study: Proof of Concept Automated Handling of Deliberate Process Deviations 127
- 4.3.3.1 Perfusion Process Deviation Analysis (Bioreactor Temperature Shift) *127*

viii Contents

4.3.3.2	Downstream Process Deviation Analysis (Viral Inactivation pH) <i>128</i>		
4.4	Summary 129		
	Acknowledgments 129		
	References 129		
	Part Three: Single Use Technologies and Perfusion		
	Technologies 131		
F	Single Lice Disconstors for Continuous Disprospersing, Challenges and		
5	Single-Use bioreactors for Continuous bioprocessing: Challenges and Outlook 123		
	Nico M.G. Oosterhuis		
51	Introduction 133		
5.2	Single Lice Departor Types 125		
53	Single-Use Reactor Types 135 Material Asposts 126		
5.5 5.4	Sensors 139		
5.5	Reactor Design 141		
551	Mass Transfer and Mixing Requirements for Continuous		
5.5.1	Processing 141		
5.6	Scale-Up Aspects 142		
5.7	Continuous Seed Train 145		
5.8	New Mixer Designs 145		
5.9	Future Outlook 146		
	References 147		
E	Two Mutually Frakling Trands Continuous Diance costing		
0	and Single Use Technologies 140		
	And Single-Use rechnologies 149 Mare Disschors, Mark Schofold, and Julia Crass		
61	Introduction 140		
6.2	Single Lice Technologica 150		
0.2	History of Single Lize Technologies 150		
6.2.1	Single Lie Unstreem Drocessing 151		
6.2.2	Single-Use Opstream Processing 151		
0.2.3	Tangantial Flow Filtration 151		
6.2.2.1	Chromotography Stong 152		
6.2.3.2	Early Skapticizm 152		
0.2.4 6.2.5	Current Trends and Euture Dradictions 152		
6.2.5	Continuous Bioprocessing 154		
6.2.1	Continuous Dioprocessing 154		
622	Continuous Opstream Processing 154		
6221	Continuous Downstream Processing 155		
6222	Langenual Flow Filtration 156		
0.3. <i>2.2</i>	Continuous Chromatography 157		
0.3.3 6 4	Concerns for Continuous Bioprocessing 158		
0.4 6 / 1	Case 1: Consume 150		
0.4.1 6 4 0	Case 1. Genzymie 137 Case 2: Marsk 160		
0.4.2	Case 2. IVIEICK 100		

Contents ix

- 6.4.3 Case 3: Bayer Technology Services 161
- 6.4.4 Comparison 162
- 6.4.5 Challenges and Solutions 163
- 6.4.6 Alternative Scenarios *164*
- 6.5 Regulatory Aspects 164
- 6.6 Adoption Rate of Single-Use and Continuous Bioprocessing 165
- 6.7 Conclusions 166 References 167
- 7 Perfusion Formats and Their Specific Medium Requirements 171

Jochen B. Sieck, Christian Schild, and Jörg von Hagen

- 7.1 Introduction 171
- 7.1.1 History of Perfusion 172
- 7.1.2 Comeback of Perfusion 172
- 7.2 Characterization of Perfusion Processes 173
- 7.2.1 Productivity of Perfusion Processes 175
- 7.2.2 Cell Retention Devices 176
- 7.2.3 Steady-State Definition 176
- 7.3 Perfusion Formats 177
- 7.3.1 Innovative Perfusion Formats 178
- 7.4 Development Strategies for Perfusion Media 179
- 7.4.1 Cell Line-Specific Requirements 181
- 7.4.2 Scale-Down Models for Perfusion Processes 181
- 7.4.3 Scale-Down Cultivation Methods 182
- 7.4.4 Examples for Perfusion Scale-Down Applications 184
- 7.5 Process Development for Perfusion Processes 185
- 7.6 Case Study 185
- 7.6.1 Material & Methods 186
- 7.6.1.1 Semicontinuous Chemostat (SCC) 187
- 7.6.1.2 Repeated Batch (RB) 187
- 7.6.1.3 Semicontinuous Perfusion (SCP) 187
- 7.6.2 Results 187
- 7.6.2.1 Determination of the Starting Cell Density 187
- 7.6.3 Scale-Down Model Comparison 188
- 7.6.4 Media Screening 189
- 7.6.5 Bioreactor Confirmation 191
- 7.7 Conclusion 192 Abbreviations 193 References 194

Part Four: Continuous Upstream Bioprocessing 201

- 8 Upstream Continuous Process Development 203 Sanjeev K. Gupta
- 8.1 Introduction 203
- 8.2 Upstream Processes in Biomanufacturing 205

- **x** Contents
 - 8.2.1 Upstream Operating Modes 206
 - 8.2.1.1 Fed-Batch Process 206
 - 8.2.1.2 Continuous/Perfusion Process 207
 - 8.3 The Upstream Continuous/Perfusion Process 207
 - 8.3.1 Upstream Process-Type Selection 209
 - 8.3.2 Component of Continuous Upstream and Downstream Processes 209
 - 8.3.2.1 Upstream Components: Stainless Steel and Single-Use (Su) 209
 - 8.3.2.2 Downstream Components: Stainless Steel and Single-Use (Su) 209
 - 8.3.3 Cell Retention Devices Used in Perfusion Process 210
 - 8.3.3.1 Spin Filters 210
 - 8.3.3.2 The ATF System 210
 - 8.3.3.3 Biosep Acoustic Perfusion System 212
 - 8.3.3.4 TFF Cell Retention Device 213
 - 8.4 Manufacturing Scale-Up Challenges 214
 - 8.4.1 Process Complexity and Control 214
 - 8.4.2 Cell Line Stability 215
 - 8.4.3 Validation 215
 - 8.5 Single-Use Technologies: A Paradigm Change 215
 - 8.5.1 Application of SUBs in Continuous Processing 218
 - 8.5.2 Single-Use Continuous Bioproduction 218
 - 8.5.3 Single-Use Perfusion Bioreactors 219
 - 8.5.3.1 Type of Single-Use Bioreactors for Perfusion Culture 219
 - 8.5.4 Single-Use Accessories Supporting Perfusion Culture 220
 - 8.5.4.1 Hollow Fiber Media Exchange 220
 - 8.5.4.2 Continuous Flow Centrifugation 220
 - 8.5.4.3 Acoustic Wave Separation 220
 - 8.5.4.4 Spin filters 220
 - 8.6 FDA Supports Continuous Processing 221
 - 8.7 Making the Switch from Batch/Fed-Batch to Continuous Processing 222
 - 8.8 Costs and Benefits of Continuous Manufacturing 222
 - 8.9 Costs of Adoption 223
 - 8.10 Continuous Downstream Processing 223
 - 8.11 Integrated Continuous Manufacturing 224
 - 8.12 Concluding Remark 227 Acknowledgment 228 References 228

9 Study of Cells in the Steady-State Growth Space 233

Sten Erm, Kristo Abner, Andrus Seiman, Kaarel Adamberg, and Raivo Vilu

- 9.1 Introduction 233
- 9.1.1 On Physiological State of Cells: Steady-State Growth Space Analysis 234
- 9.1.2 Challenge of Comprehensive Quantitative Steady-State Growth Space Analysis (SSGSA) 236
- 9.1.3 Chemostat Culture A Classical Tool for SSGSA 236

- 9.2 Advanced Continuous Cultivation Methods Changestats 237
- 9.2.1 Accelerostat (A-stat) 237
- 9.2.2 Family of Changestats A Set of Flexible Tools for Scanning Steady-State Growth Space 240
- 9.3 Review of the Results Obtained Using the Changestats 242
- 9.3.1 Acetate Overflow Metabolism in E. Coli 242
- 9.3.2 A-Stat in Study of Physiology of Yeast 243
- 9.3.3 Integration of A-Stat with High-Throughput Omics Methods and Modeling 243
- 9.3.4 A-Stat in Bioprocess Development 243
- 9.3.5 Deceleration-stat (De-stat) 244
- 9.3.6 Dilution Rate Stat (D-Stat) 244
- 9.3.7 Auxoaccelerostats 245
- 9.3.8 Adaptastat 246
- 9.4 SSGSA Using Parallel-Sequential Cultivations 247
- 9.5 Modeling in Steady-State Growth Space Analysis 248 References 250

Part Five: Continuous Downstream Bioprocessing 259

10 Continuous Downstream Processing for Production of Biotech Therapeutics 261

Anurag S. Rathore, Nikhil Kateja, and Harshit Agarwal

- 10.1 Introduction 261
- 10.2 Continuous Manufacturing Technologies for Downstream Processing 262
- 10.2.1 Continuous Cell Lysis 262
- 10.2.2 Continuous Centrifugation 263
- 10.2.3 Continuous Refolding 264
- 10.2.4 Continuous Precipitation 267
- 10.2.5 Continuous Chromatography 267
- 10.2.6 Continuous Extraction 271
- 10.2.7 Continuous Filtration 272
- 10.3 Continuous Process Development 274
- 10.4 Case Studies Related to Continuous Manufacturing 276
- 10.5 Summary 279 References 279
- 11 Evolving Needs For Viral Safety Strategies in Continuous Monoclonal Antibody Bioproduction 289 Andrew Clutterbuck, Michael A. Cunningham, Cedric Geyer, Paul Genest, Mathilde Bourguignat, and Helge Berg
- 11.1 Introduction 289
- 11.1.1 Current Regulations and Practices 293
- 11.1.2 Evolving Needs: Process versus Regulatory 294
- 11.1.3 Current Technology Landscape 295
- 11.2 Batch versus Continuous: Potential Impacts on Virus Safety 297

xii Contents

- 11.2.1 Raw Material Safety/Testing 299
- 11.2.2 Upstream and Bioreactor Safety 301
- 11.2.3 Downstream Virus Removal Strategies 304
- 11.2.3.1 Viral Reduction by Normal Flow Filtration (NFF) 304
- 11.2.3.2 Chemical Inactivation (Low pH or Solvent Detergent) 308
- 11.2.3.3 Chromatography 311
- 11.2.3.4 Other Techniques 312
- 11.3 Validation of Viral Reduction Steps in Continuous Manufacturing Processes 313
- 11.3.1 Protein A Capture Chromatography 314
- 11.3.2 Chemical Inactivation (Low pH/Solvent Detergent) 315
- 11.3.3 Intermediate and Polishing Chromatography 315
- 11.3.4 Viral Reduction Filtration 316
- 11.4 Conclusion 318 References 319

Part Six: Continuous Chromatography 321

- 12 Multicolumn Continuous Chromatography: Understanding this Enabling Technology 323 Kathleen Mihlbachler
- 12.1 Introduction 323
- 12.2 Modes of Chromatography 326
- 12.3 Interaction Mechanisms Used in Chromatographic Systems 328
- 12.4 Batch Chromatography 330
- 12.5 Semicontinuous and Continuous Batch Chromatography 331
- 12.5.1 Single Column 331
- 12.5.2 Multicolumn Parallel Operation 333
- 12.5.3 Multicolumn Parallel and Interconnected Operation 337
- 12.6 Multicolumn, Countercurrent, Continuous Chromatography 340
- 12.6.1 Implementing Traditional SMB Technology 341
- 12.6.2 SMB Technology for Biomolecules 343
- 12.6.3 Additional Examples of SMB Purifications 349
- 12.7 Risk Assessment of Continuous Chromatography 353
- 12.8 Process Design of Continuous Capture Step 357
- 12.9 Conclusion 360 References 361
- 13Continuous Chromatography as a Fully Integrated Process in
Continuous Biomanufacturing369

Steffen Zobel-Roos, Holger Thiess, Petra Gronemeyer, Reinhard Ditz, and Jochen Strube

- 13.1 Introduction 369
- 13.2 Continuous Chromatography 370
- 13.2.1 SMB 370
- 13.2.2 Serial Multicolumn Continuous Chromatography 377

- 13.2.3 Continuous Countercurrent Multicolumn Gradient Chromatography 378
- 13.2.4 Integrated Countercurrent Chromatography 379
- 13.3 Conclusion and Outlook 386 Symbols 388 References 389
- **14 Continuous Chromatography in Biomanufacturing** 393 Thomas Müller-Späth and Massimo Morbidelli
- 14.1 Introduction to Continuous Chromatography 393
- 14.2 Introduction to Manufacturing Aspects of
 - Chromatography 396
- 14.3 Trade-Offs in Batch Chromatography 399
- 14.4 Capture Applications 400
- 14.4.1 Introduction 400
- 14.4.2 Process Principle 403
- 14.4.3 Application Examples 405
- 14.5 Polishing Applications 406
- 14.5.1 Introduction 406
- 14.5.2 MCSGP (Multicolumn Countercurrent Solvent Gradient Purification) Principle 407
- 14.5.3 MCSGP (Multicolumn Countercurrent Solvent Gradient Purification) Process Design 409
- 14.5.4 MCSGP (Multicolumn Countercurrent Solvent Gradient Purification) Case Study 412
- 14.6 Discovery and Development applications 414
- 14.7 Scale-Up of Multicolumn Countercurrent Chromatography Processes *416*
- 14.8 Multicolumn Countercurrent Chromatography as Replacement for Batch Chromatography Unit Operations *417*
- 14.9 Multicolumn Countercurrent Chromatography and Continuous Upstream *419*
- 14.10 Regulatory Aspects and Control of Multicolumn Countercurrent Processes 419 References 421
- 15 Single-Pass Tangential Flow Filtration (SPTFF) in Continuous Biomanufacturing 423 Andrew Clutterbuck, Paul Beckett, Renato Lorenzi, Frederic Sengler, Torsten Bisschop, and Josselyn Haas
 15.1 Introduction 423
 15.2 Tangential Flow Filtration in Bioproduction 426
 15.2.1 Batch versus Single-Pass Tangential Flow Filtration 426
- 15.2.2 Membrane Type and Format for TFF Applications 426
- 15.2.3 Single-Pass Tangential Flow Filtration (SPTFF) 428
- 15.2.4 Process Design 430
- 15.2.5 Laboratory-Scale Process Development Example 438

xiv Contents

15.2.6	Consideration on Equipment Configuration and Requirements	442
15.3	Validation 445	
15.3.1	Key Validation Considerations between Batch and Continuous	
	Processing 445	

- 15.3.2 Validation of Single-Pass TFF 449
- 15.4 Conclusion 453 References 453

Part Seven: Integration of Upstream and Downstream 457

16	Design of Integrated Continuous Processes for High-Quality			
	Biotherapeutics	459		

Fabian Steinebach, Daniel Karst, and Massimo Morbidelli

- 16.1 Introduction 459
- 16.2 Perfusion Cell Culture Development 463
- Objectives and Requirements 463 16.2.1
- Bioreactor Setup 463 16.2.2
- 16.2.3 Physical Bioreactor Characterization 464
- 16.3 Continuous Capture Development 466
- Objectives and Requirements 466 16.3.1
- 16.3.2 Continuous Two-Column Capture Process 467
- 16.3.3 Process Performance 468
- 16.3.4 Process Control 469
- 16.4 Operation of the Continuous Integrated Process 470
- Bioreactor Operation 470 16.4.1
- Cell Growth 470 16.4.2
- 16.4.3 Monoclonal Antibody Production 471
- 16.4.4 Monoclonal Antibody Capture 472
- 16.4.5 Process Performance 473
- Product Quality 474 16.4.6
- 16.5 Conclusion 476 Acknowledgment 477 References 477
- 17 Integration of Upstream and Downstream in Continuous Biomanufacturing 481

Petra Gronemeyer, Holger Thiess, Steffen Zobel-Roos, Reinhard Ditz, and Jochen Strube

- 17.1 Introduction 481
- Background on Upstream Development in Continuous 17.2 Manufacturing 483
- Background on Downstream Development in Continuous mAb 17.3 Manufacturing 484
- Challenges in Process Development 485 17.4
- Impact of Changing Titers and Impurities on Cost 17.4.1 Structures 485

- 17.4.2 Impurities as Critical Parameters in Process Development 487
- 17.4.3 Host Cell Proteins as Main Problem in Process Development 488
- 17.4.4 Regulatory Aspects 490
- 17.5 Trends and Integration Approaches 490
- 17.6 Methodical Approach of Integrating USP and DSP Regarding Impurity Processing 492
- 17.6.1 Case Study: Influence of Media Components on Impurity Production *494*
- 17.6.2 Case Study: Influence of Harvest Operations on Impurity Production 495
- 17.6.3 Nonchromatographic Continuous DSP Operation 497
- 17.6.3.1 ATPS 498
- 17.6.3.2 Precipitation 499
- 17.6.3.3 One Step Toward a Chromatography Free Purification Process 500
- 17.7 Conclusion and Outlook 500 References 501

Part Eight: Quality, Validation, and Regulatory Aspects 511

18	Quality Control and Re	gulatory Aspects for Continuous
	Biomanufacturing 51	3
	Guillermina Forno and F	duardo Ortí

- 18.1 Introduction 513
- 18.2 FDA Support for Continuous Manufacturing 513
- 18.3 PAT as a Facilitator for Continuous Manufacturing Implementation *514*
- 18.4 PAT Applications in the Pharmaceutical Industry 516
- 18.5 Process Validation for Continuous Manufacturing Processes 519
- 18.6 Regulatory Documents Related to Process Validation 520
- 18.7 ICH 520
- 18.8 FDA 520
- 18.9 EMA 521
- 18.10 ASTM 521
- 18.11 Special Considerations for Continuous Manufacturing Process Validation 521
- 18.12 Scale-Down for Continuous Bioprocessing 524
- 18.13 Impact of Single-Use Systems in Process Validation 526
- 18.14 Batch and Lot Definition 527
- 18.15 Conclusion 528 References 528
- **19** Continuous Validation For Continuous Processing 533

Steven S. Kuwahara

19.1 Quality Management 533

- 19.2 Regulatory Considerations 534
- 19.3 Setting Specifications 534
- 19.4 Sequence of Events 535
- 19.5 Verification of Validated States 536
- 19.6 Choice of Test Methods 536
- 19.7 Types of Monitoring 536
- 19.8 Process Stream Analyzers 538
- 19.9 Validation/Qualification of Process Stream Analyzers 538
- 19.10 Control Charting 540
- 19.11 The Moving Range Chart 541
- 19.12 Continuous Validation 541
- 19.13 Choosing Other Control Charts 542
- 19.14 Information Awareness 542
- 19.15 Cost Issues 543
- 19.16 Revalidations 544
- 19.17 Management and Personnel 544 References 545

20 Validation, Quality, and Regulatory Considerations in Continuous Biomanufacturing 549

- Laura Okhio-Seaman
- 20.1 Introduction 549
- 20.1.1 What is Continuous Biomanufacturing? 549
- 20.1.2 Improvement in Product Quality 550
- 20.1.3 Manufacturing Consistency 550
- 20.1.4 Efficient Facility and Personnel Utilization 550
- 20.1.5 Reduction in Capital Expenditure and Cost 550
- 20.2 Quality 551
- 20.2.1 Other Considerations in Quality 552
- 20.2.1.1 Contract Manufacturing Organizations (CMO's) 552
- 20.2.1.2 Good Manufacturing Practices (GMP) 555
- 20.2.1.3 Supply Chains 555
- 20.2.1.4 Change Management and Control 556
- 20.3 Validation 557
- 20.3.1 Validate to Eliminate! 557
- 20.3.2 Test Conditions for Extractable and Leachable Analysis 560
- 20.3.3 Test Solutions for Extractable and Leachable Analysis 561
- 20.3.4 Analytical Techniques for Leachables Analysis 561
- 20.3.5 Description of the Model Approach 562
- 20.3.6 Actual Formulation Approach 563
- 20.4 Regulatory 564
- 20.4.1 Current Regulatory References 565
- 20.5 Conclusion 566 Further Reading 566

Part Nine: Industry Perspectives 569

21 Evaluation of Continuous Downstream Processing: Industrial Perspective 571

Venkatesh Natarajan, John Pieracci, and Sanchayita Ghose

- 21.1 Biogen mAb Downstream Platform Process 571
- 21.2 Potential Platform Process Bottlenecks Pertaining to Large Scale Manufacturing 573
- 21.3 Continuous Downstream Process 573
- 21.3.1 Multicolumn Chromatography (MCC) for Continuous Capture 575
- 21.3.1.1 Background 575
- 21.3.1.2 Process Optimization 576
- 21.3.1.3 Experimental Results 577
- 21.3.2 Continuous Viral Inactivation 578
- 21.3.3 Connected Chromatography Steps 580
- 21.3.3.1 Comparison of Current and Pool-Less Process 581
- 21.3.4 Continuous UF/DF Processes 582
- 21.4 Productivity Comparison of Batch and Continuous Downstream Process 585 References 585

Index 587

List of Contributors

Kristo Abner

Competence Center of Food and Fermentation Technologies Akadeemia tee 15 12618 Tallinn Estonia

Kaarel Adamberg

Tallinn University of Technology Department of Chemistry and Biotechnology Akadeemia tee 15 12618 Tallinn Estonia

and

Competence Center of Food and Fermentation Technologies Akadeemia tee 15 12618 Tallinn Estonia

Harshit Agarwal

Indian Institute of Technology Department of Chemical Engineering Hauz Khas 110016 New Delhi India

Paul Beckett

Millipore SAS Process Solution Technologies 39 Route Industrielle de la Hardt 67124 Molsheim France

Helge Berg

Technology Management Millipore SAS 39 Route Industrielle de la Hardt 67124 Molsheim France

Torsten Bisschop

Millipore SAS Process Solution Technologies 39 Route Industrielle de la Hardt 67124 Molsheim France

Marc Bisschops

Pall Life Sciences Scientific Laboratory Services Nijverheidsweg 1 1671 GC Medemblik The Netherlands

Mathilde Bourguignat

Technology Management Millipore SAS 39 Route Industrielle de la Hardt 67124 Molsheim France xx List of Contributors

Mark Brower

Merck & Co Inc Biologics & Vaccines 2000 Galloping Hill Road Kenilworth, NJ 07033 USA

Rimenys J. Carvalho

Federal University of Rio de Janeiro COPPE Cell Culture Engineering Laboratory C.P. 68502 21941-972 Rio de Janeiro, RJ Brazil

Leda R. Castilho

Federal University of Rio de Janeiro COPPE Cell Culture Engineering Laboratory C.P. 68502 21941-972 Rio de Janeiro, RJ Brazil

Cedric Geyer

Technology Management Millipore SAS 39 Route Industrielle de la Hardt 67124 Molsheim France

Andrew Clutterbuck

Millipore SAS Process Solution Technologies 39 Route Industrielle de la Hardt 67124 Molsheim France

M. Nicolas Cruz-Bournazou

Technische Universität Berlin Department of Biotechnology Ackerstrasse 76 ACK 24 13355 Berlin Germany

Michael A. Cunningham

Technology Management EMD Millipore Corporation 290 Concord Road Billerica, MA 01821 USA

Reinhard Ditz

Clausthal University of Technology Institute for Separation and Process Technology Leibnizstr 15 38678 Clausthal-Zellerfeld Germany

Sten Erm

Tallinn University of Technology Department of Chemistry and Biotechnology Akadeemia tee 15 12618 Tallinn Estonia

and

Competence Center of Food and Fermentation Technologies Akadeemia tee 15 12618 Tallinn Estonia

Guillermina Forno

Ciudad Universitaria Cell Culture Laboratory UNL FBCB Paraje el Pozo CC 242 Santa Fe Argentina

and

Ciudad Universitaria R&D Zelltek S.A. UNL FBCB Paraje el Pozo CC 242 Santa Fe Argentina

Paul Genest

Technology Management EMD Millipore Corporation 290 Concord Road Billerica, MA 01821 USA

Sanchayita Ghose

Bristol-Myers Squibb Downstream Process Development 38 Jackson Road Danvers, MA 01923 USA

Julie Grace

Pall Life Sciences Scientific Laboratory Services 20 Walkup Drive Westborough, MA 01581 USA

Petra Gronemeyer

Clausthal University of Technology Institute for Separation and Process Technology Leibnizstr 15 38678 Clausthal-Zellerfeld Germany

Sanjeev K. Gupta

Ipca Laboratories Ltd. Advanced Biotech Lab Kandivli Industrial Estate Kandivli (west) 400067 Mumbai India

Josselyn Haas

Millipore SAS Process Solution Technologies 39 Route Industrielle de la Hardt 67124 Molsheim France

Daniel Karst

ETH Zurich Institute for Chemical and Bioengineering Department of Chemistry and Applied Biosciences Vladimir-Prelog-Weg 1 8093 Zurich Switzerland

Nikhil Kateja

Indian Institute of Technology Department of Chemical Engineering Hauz Khas 110016 New Delhi India

Steven S. Kuwahara

GXP BioTechnology LLC Tucson, AZ 85741 USA

Renato Lorenzi

Millipore SAS Process Solution Technologies 39 Route Industrielle de la Hardt 67124 Molsheim France

Kathleen Mihlbachler

Lewa Process Technologies Inc. Separations Development 8 Charlestown Street Devens, MA 01434 USA

Massimo Morbidelli

ETH Zurich Institute for Chemical and Bioengineering Department of Chemistry and Applied Biosciences Vladimir-Prelog-Weg 1 8093 Zürich Switzerland

Thomas Müller-Späth

ChromaCon AG Process Development Technoparkstrasse 1 8005 Zurich Switzerland

and

ETH Zurich Institute for Chemical and Bioengineering Department of Chemistry and Applied Biosciences Vladimir-Prelog-Weg 1 8093 Zürich Switzerland

Venkatesh Natarajan

Biogen Engineering & Technology 225 Binney Street Cambridge, MA 02142 USA

Peter Neubauer

Technische Universität Berlin Department of Biotechnology Ackerstrasse 76 ACK 24 13355 Berlin Germany

Laura Okhio-Seaman

Sartorius Stedim North America Validation Services 5 Orville Drive Bohemia, NY 11716 USA

Nico M.G. Oosterhuis

Celltainer Biotech BV Bothoekweg 9 7115AK Winterswijk The Netherlands

Eduardo Ortí

Ciudad Universitaria R&D Zelltek S.A. UNL FBCB Paraje el Pozo CC 242 Santa Fe Argentina

John Pieracci

Biogen Engineering & Technology 225 Binney Street Cambridge, MA 02142 USA

David Pollard

Merck & Co Inc Biologics & Vaccines 2000 Galloping Hill Road Kenilworth, NJ 07033 USA

Anurag S.Rathore

Indian Institute of Technology Department of Chemical Engineering Hauz Khas 110016 New Delhi India

Douglas Richardson

Merck & Co Inc Biologics & Vaccines 2000 Galloping Hill Road Kenilworth, NJ 07033 USA

Christian Schild

Merck Life Science (a business of Merck KGaA) Process Solutions Cell Culture Media R&D Frankfurter Strasse 250 64291 Darmstadt Germany

Mark Schofield

Pall Life Sciences Applications R&D 20 Walkup Drive Westborough, MA 01581 USA

Andrus Seiman

Tallinn University of Technology Department of Chemistry and Biotechnology Akadeemia tee 15 12618 Tallinn Estonia

and

Competence Center of Food and Fermentation Technologies Akadeemia tee 15 12618 Tallinn Estonia

Frederic Sengler

Millipore SAS Process Solution Technologies 39 Route Industrielle de la Hardt 67124 Molsheim France

Jochen B. Sieck

Merck Life Science (a business of Merck KGaA) Process Solutions Cell Culture Media R&D Frankfurter Strasse 250 64291 Darmstadt Germany

Fabian Steinebach

ETH Zurich Institute for Chemical and Bioengineering Department of Chemistry and Applied Biosciences Vladimir-Prelog-Weg 1 8093 Zurich Switzerland

Jochen Strube

Clausthal University of Technology Institute for Separation and Process Technology Leibnizstr 15 38678 Clausthal-Zellerfeld Germany

Holger Thiess

Clausthal University of Technology Institute for Separation and Process Technology Leibnizstr 15 38678 Clausthal-Zellerfeld Germany

Raivo Vilu

Tallinn University of Technology Department of Chemistry and Biotechnology Akadeemia tee 15 12618 Tallinn Estonia

xxiv List of Contributors

and

Competence Center of Food and Fermentation Technologies, Akadeemia tee 15 12618 Tallinn Estonia

Jörg von Hagen

Merck Life Science (a business of Merck KGaA) Process Solutions Cell Culture Media R&D Frankfurter Strasse 250 64291 Darmstadt Germany

Steffen Zobel-Roos

Clausthal University of Technology Institute for Separation and Process Technology Leibnizstr 15 38678 Clausthal-Zellerfeld Germany Part One

Overview of State-of-the-Art Technologies and Challenges

Continuous Bioprocess Development: Methods for Control and Characterization of the Biological System

3

Peter Neubauer and M. Nicolas Cruz-Bournazou

Technische Universität Berlin, Department of Biotechnology, Ackerstrasse 76, ACK 24, 13355 Berlin, Germany

1.1 Proposed Advantages of Continuous Bioprocessing

1.1.1 Introduction

The change from batch to continuous processing has led to the intensification of processes in a number of industries, including steel casting, automobile and other devices, petrochemicals, food, and pharmaceuticals. Advantages include, aside from a significant increase in volumetric productivity, reduced equipment size, steady-state operation, low cycle times, streamed process flows, and reduced capital cost.

In bioengineering, continuous processing is the standard in wastewater treatment, composting, and some bioenergy processes such as biogas and bioethanol fermentations. In contrast, most production processes run as batch type operations or more specifically fed-batch processes, which is the major production technology today.

Konstantinov and Cooney provide a definition of a continuous process as "A unit operation is continuous if it is capable of processing a continuous flow input for prolonged periods of time. A continuous unit operation has minimal internal hold volume. The output can be continuous or discretized in small packets produced in a cyclic manner." [1]. They also differentiate between full continuous processes with no or minimal hold volume in the process line or hybrid processes that contain both batch and continuous process operations.

Obviously, the push in continuous manufacturing technologies was initiated by the BioPAT initiative of the Food and Drug Administration (FDA) in 2002 and the published guidance to PAT in 2004 [2], which initially aimed at a better understanding of the connections between product quality and process conditions. This lead to the need to develop quality by design (QbD), that is, the implementation of process analytical tools over the whole developmental pipeline from early product screening over the process development in the laboratory scale and during scale up. The needs for a better understanding of the impact of process parameters on the critical quality attributes (CQA) of the respective product also increased the interest in the development and implementation of novel sensors and analytical

Continuous Biomanufacturing: Innovative Technologies and Methods, First Edition. Edited by Ganapathy Subramanian.

@ 2018 Wiley-VCH Verlag GmbH & Co. KGaA. Published 2018 by Wiley-VCH Verlag GmbH & Co. KGaA.

4 1 Continuous Bioprocess Development

tools. As a consequence, this better understanding of processes resulted in further process intensification and provided the instrumental basis to approach challenges in relation to continuous operation.

Aside the FDA initiative, there are several drivers for the increasing interest in continuous processing, not only in the pharmaceutical industry but also in the industrial (white) biotech industry. On one side, we see an increasing demand and thus also increasing production scale for industrial bioproducts (enzymes, small molecules, and bioenergy market) with a need for reduced costs for the products and increased competition. Considering that production scales are steadily growing and that a scale reduction close to factor 10 would be possible by continuous processing, plant sizes and the efficiency of bioprocesses could be increased significantly. On the other side, the opportunity of the selection of new biocatalysts and its implementation in the chemical synthesis for integrated chemoenzymatic processes (i.e., processes which combine chemical and enzymatic reactions) have to be competitive with the existing chemical processes and need to be integrated into the chemical production schemes. Here, continuous processes offer clear advantages.

In biopharma for recombinant proteins, antibodies, highly complex proteins, recombinant enzymes and blood factors, the efficiency of the cell factories, and production systems have dramatically increased during the last decade. Opportunities for high cell density processes with a higher volumetric product yield and quality, as well as the changing situation in view of the intellectual properties by the termination of many patents for important drugs with novel commercial opportunities for new biosimilars and biobetters are a strong driver in increasing the competition especially from emerging markets. In parallel, there is an increasing demand for establishing local production sites for defined regional markets, rather than having single production sites. Strict cost calculations as a developmental driver demand for smaller and effective, but also flexible production plants. This directs interest to evaluate continuous bioprocessing opportunities to minimize investments for production facilities, and thinking about parallelization rather than larger scales. Parallelization would also be an advantage in processes with longer plant cycle times [3] as, for example, cell culture-based products. A nice example that shows the opportunities in significantly decreasing operational and capital expenses by changing from conventional bioprocessing to continuous bioprocessing in the case of production on monoclonal antibodies (mAB) and other non-mAB processes is shown by Walther et al. [4].

However, despite the obvious opportunities of continuous processes there are many challenges to solve, mainly the demand for fast realization and risk minimization. Currently, it seems to be easier to transfer a batch process into production than to start a new, longer, and more expensive development of a continuous process even though it is expected to be more efficient.

These scenarios show that there is a big need in strategic methods concerning the development of continuous process strategies for either new products or to derive a continuous process from existing batch type processes. As early-phase product development can practically be only performed as batch processes, a key question in product development is how we can transfer a batch strategy to a continuous operation in a large process.